

# Isolation and Characterization of an *Aeromonas punctata* Bacteriophage

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**Abstract:** An *Aeromonas punctata* bacteriophage, named as DH1, was isolated from East Lake, Wuhan city, China. Morphologically, phage DH1 showed a typical *Myoviridae* structure consisting of an isometric head (50 nm in diameter) and a visible tail. The bacteriophage had a latent period of about 90 minutes and an average burst size of about 125 PFU•Cell<sup>-1</sup>. Restriction enzyme pattern of the bacteriophage's genome showed that the genome is a double-stranded DNA and about 34kb in size. The sequenced genomic fragments showed highly similarities to gp04 and gp16 sequence of other *Myoviridae* bacteriophages at protein level.

**Keywords:** *Myoviridae*, bacteriophage, *Aeromonas punctata*.

## 1. INTRODUCTION

*Aeromonas sp.* has been well known to cause diseases in fishes, frogs, and even humans [1-4]. Till now, only one *Aeromonas hydrophila* bacteriophage has been isolated [5] and sequenced [6]. In this study, we reported the isolation and characterization of a novel *Aeromonas punctata* bacteriophage from a natural lake.

## 2. MATERIALS AND METHODS

**Sampling:** The sampling site was located in East Lake (30°31'37.66"N, 114°22'09.18"E), Wuhan City, Hubei Province, China. Surface water samples (10-20cm depth) were collected and transported to laboratory immediately for bacteria and virus isolation during December 23, 2006 to January 8, 2007.

### 2.1. Isolation of Bacteria and Bacteriophage

A volume of 100μL water from lake was spread over solid LB media. After incubation at 37°C for 24h, separated colony was picked and purified by standard methods. The isolation of bacteriophage was performed using phage plaque method. Briefly, 30mL fresh lake water was filtered through a 0.2μm filter (PN4612, PALL). A volume of 100μL filtered water was mixed with 200μL bacterial culture in LB media and incubated at 28°C till the plaque was formed. Pure bacteriophage strain was obtained by five serial single-plaque isolations. Only one phage-host system was established from 60 purified bacterial colonies, the phage was named as DH1, and the host bacterium was designated MH1.

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### 2.2. Characterization of MH1

The genomic DNA of bacteria MH1 was extracted with phenol-chloroform-isoamylol and precipitated with ethanol. A fragment of 16S rDNA was amplified by one pair of universal primer [7] and sequenced.

### 2.3. Phage Purification

Five hundred milliliters of cell lysates (the titer of the phage was about 10<sup>9</sup>PFU•mL<sup>-1</sup>) was centrifuged at 10000 g for 30 min in Jouan KR2.5i centrifuge to remove the cell debris. Then, the supernatant was centrifuged at 100,000 g for 2.5 h in Beckman J-30I (JS-24 rotor) to concentrate the phage stock to about 2mL. The phage stock was further purified on 20%-50% sucrose density-gradient centrifugation by Beckman optima<sup>™</sup> MAX and MAX-E(MLS-50rotor). Finally, the phage stock was diluted in 3mL 20mmol/L MgCl<sub>2</sub> solution and stored at 4°C.

### 2.4. Transmission Electron Microscopy (TEM)

The purified phages were dropped onto a formvar carbon-coated copper grid (200 mesh) and negatively stained with 2% sodium phosphotungstate (pH7.0) and observed under TEM (Hitachi H-7000FA).

### 2.5. Phage DNA Extraction and Restriction Endonuclease Digestion

The nucleic acid of purified phage was extracted using protease K, followed by phenol-chloroform-isoamylol extraction and ethanol precipitation. The phage genomic DNA was digested with endonuclease *Acc I*, *BamH I* and *Sau3A I* (Takara) and load onto a 0.8% agarose gel and analyzed using Ethidium Bromide staining.

## 2.6. Cloning and Sequencing

Two *BamH* I fragments, 1.4 kbp and 1.9 kbp were purified with a gel extraction kit (D2501-01, OMEGA), then cloned to pUC19 vector using standard protocol [8]. The recombinant plasmids DNA were purified and sequenced with universal primers by Invitrogen Company.

## 2.7. Growth Cycle

Phage DH1 and exponential phase host cell MH1 were mixed at a MOI (multiplicity of infection) of 1:2000. The mixture was cultured at 28°C, and plaque assay was used to test the titer of DH1 every 30 min during 2.5 hours.

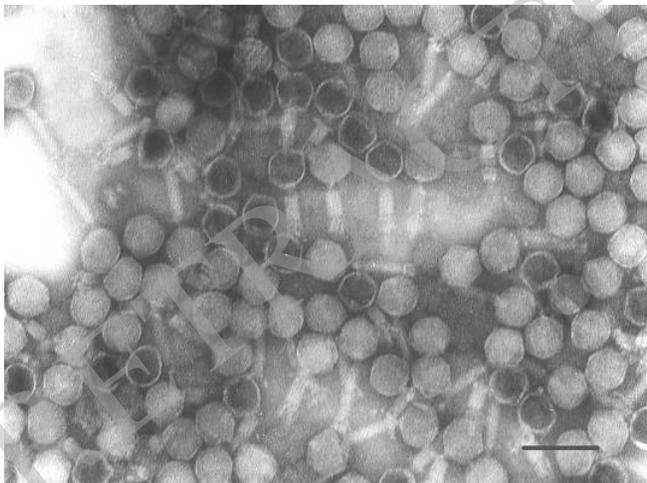
## 3. RESULTS AND DISCUSSION

### 3.1. Identification of the Host

A fragment of 196 bp of 16S rDNA was amplified and sequenced from bacteria MH1, and the sequence showed 100% identity to *Aeromonas punctata* and was submitted to GenBank (access number: EU515214).

### 3.2. Identification of the Phage

TEM analysis showed that phage DH1 had an icosahedral head (50 nm in diameter) and a visible tail (less than 100 nm in length) (Fig. 1), which is morphologically similar to but much smaller than *Aeromonas hydrophila* phages Aeh1 and Aeh2 [5].

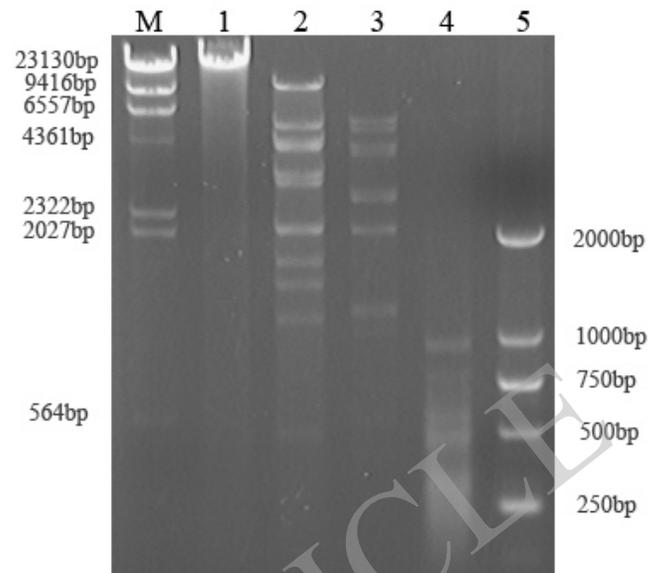


**Fig. (1).** Electron micrograph of negatively stained phage DH1.

Bar: 100nm

### 3.3. Genome Analysis of DH1

The result of enzyme digestion showed that the genome size is approximately 34±3kb (Fig. 2) and much lesser than Aeh1(230kb) [9]. The sequence of two fragments, 1.4 and 1.9kb, were submitted to GenBank (access number: EU515215 and EU515216) and showed 42% and 50% identities to gp04 and gp16 of a *Myoviridae* bacteriophage Bcep43, respectively [10].



**Fig. (2).** Restriction enzyme digested patterns of DH1 genome DNA in 0.8% agarose gel electrophoresis.

M:  $\lambda$  phage DNA digested by HindIII, 1: phage genome DNA without digestion, 2: Acc I, 3: BamH I, 4: Sau3A I, 5:DL2000 marker.

### 3.4. Growth Cycle

The one-step growth curves of DH1 indicated that the latent periods of phage DH1 were 90 min, which was much longer than *Aeromonas hydrophila* phages Aeh1 and Aeh2 [5]. The average burst sizes of phage DH1 were about 125 PFU•Cell<sup>-1</sup>, which was also bigger than Aeh1 and Aeh2 [5].

The majority of *Aeromonas sp.* described to date are from soil and water and known to be pathogenic to cold-blooded animals. Some studies have demonstrated that the presence of *A. sp.* in drinking water is a potential risk, since some strains can produce a wide range of virulence factors [2, 4]. Thus, by studying the interaction of the phage-host system, *A. sp.* phage may be significant for controlling the risk [11].

## CONCLUSION

*A. punctata* bacteriophage DH1 was a typical Myoviridae bacteriophage, and its genome is a 34kb double-stranded DNA. The sequenced genomic fragments showed highly similarities to other Myoviridae bacteriophages.

## CONFLICT OF INTEREST

The authors confirm that this article content has no conflict of interest.

## ACKNOWLEDGEMENTS

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